

## TRITERPENOIDS FROM THE FEMALE AND MALE FLOWERS OF *ALNUS SIEBOLDIANA*

FUKIKO SAKAMURA\*, SHINJI OHTA†, TADASHI AOKI† and TAKAYUKI SUGA†‡

\*Department of Food, Oshimo Women's College, Gion-cho, Asaminami-ku, Hiroshima 731-01, Japan; †Department of Chemistry, Faculty of Science, Hiroshima University, Higashisenda-machi, Naka-ku, Hiroshima 730, Japan

(Revised received 15 April 1985)

**Key Word Index**—*Alnus sieboldiana*; Betulaceae; female flowers; male flowers; C<sub>31</sub>-secodammarane-type triterpenoids.

**Abstract**—The major triterpenoids in both the female and male flowers of *Alnus sieboldiana* are a C<sub>31</sub>-secodammarane-type, alnustic acid, and its 12-*O*- $\alpha$ -L-arabinofuranoside and 12-*O*- $\beta$ -D-xylopyranoside.

Previous studies on the chemical constituents of *Alnus sieboldiana* Matsum. (Japanese name: Ohba-yashabushi) have shown the presence of flavonoids in the pollen [1, 2] and hydrocarbons [3], flavonoids and stilbenes [4], diarylheptanoids [5, 6] and a C<sub>31</sub>-secodammarane-type triterpenoid, such as alnustic acid [7], in the male flowers. Alnustic acid and its monoglycosides have been also isolated from the female flowers of *A. serrulatoides* Call. [8] and the male flowers of *A. pendula* Matsum. [9].

We examined the flowers of *A. sieboldiana*, and found the presence of alnuserrudiolone (1) and alnustic acid (2) and its 12-*O*- $\alpha$ -L-arabinofuranoside (3), 12-*O*- $\beta$ -D-xylopyranoside (4) and 12-*O*- $\beta$ -D-glucopyranoside (5) in the female flowers and the presence of the glycosides 3 and 4, in addition to alnustic acid (2), in the male flowers. Table 1 shows the relative quantities, the triterpenoids being numbered in the order of increasing polarity on TLC.

Alnustic acid (2) and its 12-*O*- $\alpha$ -L-arabinofuranoside (3)

and 12-*O*- $\beta$ -D-xylopyranoside (4) predominate in the female and male flowers (Table 1). Alnuserrudiolone (1) [10], a C<sub>31</sub>-dammarane-type triterpenoid, and alnustic acid 12-*O*- $\beta$ -D-glucopyranoside (5) were present in only the female but not the male flowers. Compounds 2–5 have been also found to be present in the female flowers of *A. serrulatoides* [8] and the male flowers of *A. pendula* [9]. Although various triterpenoids had been isolated from the leaves, barks, cortices and twigs of many species of Betulaceae [11–14], no C<sub>31</sub>-secodammarane-type has yet been isolated. Thus, the present findings and our previous results [8, 9] indicate that the C<sub>31</sub>-secodammarane-type triterpenoid is a characteristic compound in the flowers of this genus. Investigations on triterpenoids in the flowers of other *Alnus* species are now in progress.

### EXPERIMENTAL

All the equipment used has been described previously [9]. Analytical TLC was performed on silica gel (Merck No. 5715).

‡To whom correspondence should be addressed.

Table 1. Triterpenoids in the female and male flowers of *A. sieboldiana*

| Compounds*  | Relative amount† |      | R <sub>f</sub> ‡ |      |      |
|---|------------------|------|------------------|------|------|
|   | Female           | Male | (a)              | (b)  | (c)  |
| Alnuserrudiolone (1)                                | +                | —    | 0.19             | 0.81 |      |
| Alnustic acid (2)                                   | +++              | +++  | 0.03             | 0.59 | 0.91 |
| Al 12- <i>O</i> - $\alpha$ -L-arabinofuranoside (3) | ++++             | ++++ |                  | 0.29 | 0.74 |
| Al 12- <i>O</i> - $\beta$ -D-xylopyranoside (4)     | +++              | +++  |                  | 0.19 | 0.68 |
| Al 12- <i>O</i> - $\beta$ -D-glucopyranoside (5)    | ++               | —    |                  |      | 0.59 |

\*Al = Alnustic acid.

†Estimated by visual examination on TLC plates.

‡Solvent systems: (a) EtOAc–hexane (2:3); (b) CHCl<sub>3</sub>–MeOH (9:1) and (c) CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (40:10:1).

Compounds were visualized as coloured spots by spraying with vanillin-H<sub>2</sub>SO<sub>4</sub> (1:134, w/w) and then by heating on a hot-plate and/or by spraying with 0.3% *p*-Bromocresol Green soln in H<sub>2</sub>O-MeOH (1:4) adjusted to pH 8.0.

**Extraction and isolation.** The female flowers (483 g) of *A. sieboldiana*, growing wild on a mountain in the suburbs of Hiroshima City, were collected in the middle of March (ca 1 week after the flowering of the male flowers). The female flowers, after they had been minced mechanically, were immersed in Me<sub>2</sub>CO at room temp for 10 months. Removal of the solvent from the Me<sub>2</sub>CO soln gave a brown, viscous oily substance (20.0 g). This oily substance was subjected to centrifugal chromatography on silica gel [Merck 60; 5 mm × 30 cm (diam)] by means of gradient elution with CHCl<sub>3</sub>-MeOH as eluant (MeOH increasing from 0 to 100%), followed by purification with prep. TLC to give compounds 1-5. The male flowers (5.0 kg) were also collected just after flowering from the same trees and extracted as described for the case of the female flowers to give an Me<sub>2</sub>CO extract (130 g), which showed the presence of three triterpenoids 2-4 on TLC.

**Identification of compounds.** The triterpenoids isolated from the female flowers were alnuserrudiolone (1, 5 mg, an amorphous solid) and alnustic acid (2, 96 mg, mp 154-156°) and its 12-*O*-α-L-arabinofuranoside (3, 144 mg, an amorphous solid), 12-*O*-β-D-xylopyranoside (4, 101 mg, an amorphous solid) and 12-*O*-β-D-glucopyranoside (5, 45 mg, an amorphous solid). All these compounds showed identical behaviour on TLC, physico-chemical data and spectra as those of known samples.

Compounds 2-4 in the male flowers were identified by co-TLC with those isolated from the female flowers.

#### REFERENCES

1. Ohmoto, T., Nikaido, T., Nozaki, T. and Ikuse, M. (1977) *Yakugaku Zasshi* **97**, 176.
2. Ohmoto, T. and Nikaido, T. (1980) *Shoyakugaku Zasshi* **34**, 316.
3. Hayashi, S., Sakamura, F., Asakawa, Y. and Matsuura, T. (1970) *Nippon Nogekagaku Kaishi* **44**, 218.
4. Asakawa, Y. (1971) *Bull. Chem. Soc. Jpn.* **44**, 2761.
5. Asakawa, Y. (1972) *Bull. Chem. Soc. Jpn.* **45**, 1794.
6. Suga, T., Ohta, S., Aoki, T. and Hirata, T. (1983) *Bull. Chem. Soc. Jpn.* **56**, 3353.
7. Suga, T., Aoki, T., Hirata, T., Aoki, K. and Asakawa, Y. (1979) *Bull. Chem. Soc. Jpn.* **52**, 1698.
8. Aoki, T., Ohta, S., Aratani, S., Hirata, T. and Suga, T. (1982) *J. Chem. Soc. Perkin Trans. 1*, 1399.
9. Suga, T., Aoki, T., Kawada, Y., Ohta, S. and Ohta, E. (1984) *Phytochemistry* **23**, 1297.
10. Hirata, T., Aoki, T. and Suga, T. (1981) *Bull. Chem. Soc. Jpn.* **54**, 3059.
11. Kulshreshtha, M. J., Kulshreshtha, D. K. and Rastogi, R. P. (1972) *Phytochemistry* **11**, 2369.
12. Pant, P. and Rastogi, R. P. (1979) *Phytochemistry* **18**, 1095.
13. Das, M. C. and Mahato, S. B. (1983) *Phytochemistry* **22**, 1071.
14. Boar, R. B. (1984) *Nat. Prod. Rep.* **53**.